

# The Journal of Biomedical Research

#### Glioblastoma multiforme: Diagnosis, treatment, and invasion

Li Jiawei, Feng Lili, Lu Yingmei

Cite this article as:

Li Jiawei, Feng Lili, Lu Yingmei. Glioblastoma multiforme: Diagnosis, treatment, and invasion[J]. *Journal of Biomedical Research*, 2023, 37(1): 47–58. doi: 10.7555/JBR.36.20220156

View online: https://doi.org/10.7555/JBR.36.20220156

#### Articles you may be interested in

Herpes zoster in breast cancer: a case report

The Journal of Biomedical Research. 2017, 31(4): 370 https://doi.org/10.7555/JBR.31.20170009

Exposure to environmental bisphenol A inhibits HTR-8/SVneo cell migration and invasion

The Journal of Biomedical Research. 2020, 34(5): 369 https://doi.org/10.7555/JBR.34.20200013

Cholesterol crystal embolization following plaque rupture: a systemic disease with unusual features The Journal of Biomedical Research. 2017, 31(2): 82 https://doi.org/10.7555/JBR.31.20160100

Medical management of COVID-19 clinic

The Journal of Biomedical Research. 2020, 34(6): 416 https://doi.org/10.7555/JBR.34.20200118

Spindle cell carcinoma of the mandible: a case report

The Journal of Biomedical Research. 2017, 31(3): 273 https://doi.org/10.7555/JBR.30.20140061

Evaluation of atherosclerotic lesions in cholesterol-fed mice during treatment with paclitaxel in lipid nanoparticles: a magnetic resonance imaging study

The Journal of Biomedical Research. 2017, 31(2): 116 https://doi.org/10.7555/JBR.31.20160123



Available online at www.jbr-pub.org.cn

Open Access at PubMed Central

The Journal of Biomedical Research, 2023 37(1): 47–58

JBR

Review Article

### Glioblastoma multiforme: Diagnosis, treatment, and invasion

Jiawei Li<sup>1,2</sup>, Lili Feng<sup>3,⊠</sup>, Yingmei Lu<sup>1,⊠</sup>

<sup>1</sup>Department of Physiology, School of Basic Medical Sciences, Nanjing Medical University, Nanjing, Jiangsu 211166, China;

<sup>2</sup>The First Clinical Medical College of Nanjing Medical University, Nanjing, Jiangsu 211166, China; <sup>3</sup>Key Laboratory of Cardiovascular & Cerebrovascular Medicine, Drug Target and Drug Discovery Center, School of Pharmacy, Nanjing Medical University, Nanjing, Jiangsu 211166, China.

#### Abstract

Glioblastoma multiforme (GBM) is an essentially incurable brain tumor, which has been explored for approximately a century. Nowadays, surgical resection, chemotherapy, and radiation therapy are still the standardized therapeutic options. However, due to the intrinsic invasion and metastasis features and the resistance to chemotherapy, the survival rate of glioblastoma patients remains unsatisfactory. To improve the current situation, much more research is needed to provide comprehensive knowledge of GBM. In this review, we summarize the latest updates on GBM treatment and invasion. Firstly, we review the traditional and emerging therapies that have been used for GBM treatment. Given the limited efficiency of these therapies, we further discuss the role of invasion in GBM recurrence and progression, and present current research progress on the mode and mechanisms of GBM invasion.

Keywords: glioblastoma multiforme, diagnosis, treatment, patterns of invasion, invasion mechanism

#### Introduction

Glioblastoma multiforme (GBM) is the most aggressive malignant brain tumor with a high incidence rate and a low survival rate. It accounts for approximately 14.7% of all central nervous system tumors (CNSTs), and 56.5% of gliomas<sup>[1]</sup>. The overall incidence rate in the United States is 4.23%, while Asians/Pacific islanders have a relatively low incidence rate of 2.00%<sup>[2]</sup>. With a bleak prognosis due to the high aggression and recurrence rate, the median survival time is 12 months in all GBM cases, and the average 2-year and 5-year survival rates were 21.3% and 13.8% respectively<sup>[3]</sup>. In China, the burden of CNSTs is nonnegligible with a large number of affected individuals, and the overall CNST incident cases have increased by 106.52% within the past three decades<sup>[4]</sup>.

Primary GBM, accounting for 94.7% of GBM, has a mean age at occurrence between 59 and 62, while a secondary GBM, which is rarer, occurs at a relatively young age. The two types of glioblastomas both

#### lufx@njmu.edu.cn.

<sup>&</sup>lt;sup>™</sup>Corresponding authors: Lili Feng, Key Laboratory of Cardiovascular & Cerebrovascular Medicine, Drug Target and Drug Discovery Center, School of Pharmacy, Nanjing Medical University, 101 Longmian Avenue, Jiangning District, Nanjing, Jiangsu 211166, China. Tel: +86-25-86868462, E-mail: fenglilinjmu@njmu.edu.cn; Yingmei Lu, Department of Physiology, School of Basic Medical Sciences, Nanjing Medical University, 101 Longmian Avenue, Jiangning District, Nanjing, Jiangsu 211166, China. Tel: +86-25-86868462, E-mail:

Received: 13 July 2022; Revised: 22 August 2022; Accepted: 30 August 2022; Published online: 28 October 2022

CLC number: R739.41, Document code: A

The authors reported no conflict of interests.

This is an open access article under the Creative Commons Attribution (CC BY 4.0) license, which permits others to distribute, remix, adapt and build upon this work, for commercial use, provided the original work is properly cited.

originate from glial progenitor cells or neural stem cells, but the secondary GBM derives from astrocytoma<sup>[5]</sup>. Normally, patients are treated with combined therapies, including chemotherapy, surgery, or radiotherapy, aiming to ameliorate the frustrating survival rate. Nevertheless, the intricate infiltration nature and heterogeneity of GBM hinder the complete eradication of the tumor, thus contributing to a high recurrence rate<sup>[6]</sup>.

GBM invasion induced by the diffuse infiltration and invasion margin has caught the attention from clinic doctors, pathologists, and pharmacologists for a long time. GBM cells invade following the Scherer structure, which was named after a German pathologist who defined it. These cells infiltrate along existing brain structures like brain parenchyma, blood vessels, white matter tracts, and subpial spaces. Despite their preference for the white matter, they migrate fastest alongside blood vessels, mostly unidirectionally, and in a helical movement<sup>[7]</sup>.

Among various subtypes of glioblastomas, the mesenchymal subtype is the most liable to invade. Other phenotypes may also transit into the mesenchymal subtype through the mesenchymal transition<sup>[8]</sup>. Various pathways have been shown to contribute to GBM invasion. In addition, the microenvironment also provides structural support for GBM cells and may act as a guiding scaffold in the process of invasion.

In this review, we take a glance at different methods

of treatment and diagnosis, including the common practice and some future prospects. When further exploring the poor treatment outcomes and prognosis, we found the key contribution of invasion. So, we discuss the current knowledge of the mode and dynamics of invasion, and depict a holistic picture of mechanisms whereby GBM cells invade. Based on these findings, we hope to provide more comprehensive knowledge about GBM for researchers to develop more effective treatment options.

#### Diagnosis

Techniques employed in the diagnosis of GBM comprise invasive and non-invasive ones, as shown in Fig. 1A. Contrast-enhanced magnetic resonance imaging (MRI) is the most widely used non-invasive technique<sup>[9]</sup>. For highly specific imaging, positron emission tomography (PET) can be considered, which is recommended for the diagnosis of level III/IV glioblastoma. Another innovative option is "immunotargeted imaging" in which the high targetspecific antibodies combine with the given tumor cell surface target, and PET is subsequently used for imaging. This process brings the real-time monitoring The collaboration to the reality. of immunohistochemistry and PET (immuno-PET) is thus called a "virtual biopsy"<sup>[10]</sup>.

For a definitive diagnosis, histopathological examinations are necessary, which requires tumor



*Fig. 1* Glioblastoma multiforme diagnosis and treatment. A: The current strategy for GBM diagnosis includes the application of imaging technology (MRI or PET), histopathological exams following the tumor resection, and molecular diagnostics testing for the mutation status. B: Traditional treatment of GBM comprises surgery, RT, and chemotherapy primarily with TMZ. TTF therapy, immunotherapy, and stem cell-based therapy are emerging therapies possibly of significance in the future, especially for recurrent GBM. GBM: glioblastoma multiforme; MRI: magnetic resonance imaging; PET: positron emission tomography; RT: radiation therapy; TMZ: temozolomide; TTF: tumor treating fields.

resection. If metastatic GBM is suspected, fine needle aspiration cytology is relatively more reliable, especially in the study of extra-cranial metastases to the parotid gland<sup>[11]</sup>. With the discovery of more circulating biomarkers, the liquid biopsy may become a trending auxiliary examination in the future. Serum, plasma, and cerebrospinal fluid can be sampled and analyzed for biomarkers like ctDNA, miRNA, proteins, and exosomes. Tumor cells disseminated into the fluid might also be found in a liquid biopsy<sup>[12]</sup>.

To predict the invasiveness and the prognosis of GBM, testing for the mutation status of biomarkers is required. One of the most important biomarkers is isocitrate dehydrogenase 1 and 2 (IDH1/IDH2), whose mutations have been used for GBM classification and prognosis prediction<sup>[9]</sup>. Chromosome 1p and 19q codeletion, mutations in ATRX, telomerase reverse transcriptase (TERT) promoter, tumor protein p53 (TP53), and B-raf proto-oncogene as well as serine/threonine kinase (BRAF) V600E mutation are also supportive for GBM diagnosis<sup>[13]</sup>. Moreover, O<sup>6</sup>methylguanine-DNA methyltransferase (MGMT) promoter methylation is employed to guide GBM treatment<sup>[14]</sup>. Other crucial mutations occurring in phosphatase and tensin homolog (PTEN), and variants of histone 3, the epidermal growth factor receptor (EGFR), Ki-67, and glial fibrillary acidic protein (GFAP) are reported to accelerate the progression of GBM. In addition, their mutational landscape potentially contributes to GBM subclassification and is of prognostic value, yet none of which are used as molecular biomarkers in clinical practice<sup>[15-19]</sup>. To find the much more specific biomarkers for GBM diagnosis apart from IDH, 1p/19q, and MGMT, further investigations are needed.

#### Treatment

Traditional treatments are the standard approach used for newly diagnosed GBM patients. Surgery is the first choice of the standardized GBM therapies. Maximal safe surgical resection is recommended for cases that are not suitable for total eradication, like invasive or metastatic GBM<sup>[9]</sup>. Depending on the extent of tumor resection, surgical options are divided into four types, including gross total resection (GTR), subtotal resection (SR), partial resection (PR), and biopsy<sup>[20]</sup>. A second or a third surgery might be needed in some cases, but the outcome is unsatisfactory. Patients who have received surgeries may suffer from seizures and focal neurological deficits. Headache, nausea, stupor, and unconsciousness due to intracranial hypertension are also the common side effects<sup>[21]</sup>.

As an alternative therapy, RT has shown clear advantages due to its non-invasiveness. 50 to 60 Gy is suitable for the most cases, which can eliminate microscopic lesions after tumor resection<sup>[22]</sup>, and with the application of imaging techniques, including MRI, the radiation can be limited effectively to a local extent<sup>[23]</sup>. The 3D RT with a portal imaging is the recommended technique for GBM patients. For those under the age of 70 or in good general health, RT beginning within four to six weeks after surgery or even earlier, in combination with chemotherapy, is the ideal choice. An accelerated hypofractionated RT regimen is applicable to patients over the age of 70 years and those in poor general health<sup>[24]</sup>. However, tumor relapse is still inevitable due to tumor cellmicroenvironment-mediated intrinsic or tumor resistance to RT<sup>[25]</sup>.

In addition to surgical treatment and RT, the standard postoperative care also include chemotherapy. Only three chemotherapeutic agents are approved by the Food and Drug Administration (FDA) now. The first class is nitrosoureas, including carmustine and lomustine, used for GBM chemotherapy over 40 years ago<sup>[26]</sup>. However, due to liver and kidney toxicities, they are mostly abandoned in treatment<sup>[27]</sup>. Implantable carmustine pumps are still used for the local delivery of medicine in the resection cavity to improve the survival rate of both the newly diagnosed and recurrent GBM<sup>[28]</sup>. The second approved agent is temozolomide (TMZ), which is commonly used in treating newly diagnosed malignant brain tumors. It can cross the blood-brain barrier disrupt DNA replication, and cause quickly, modification and cross-linking of DNA, which mostly results in apoptosis of rapidly dividing cells located in the brain<sup>[29]</sup>. The biggest problem of TMZ is that patients were susceptible to resistance. One of the main contributors to TMZ resistance is MGMT, which repairs TMZ-induced DNA alkylation. Base excision repair and autophagy are other suspected contributors<sup>[30]</sup>. New methods have been adopted to overcome TMZ resistance. As the first anti-angiogenic agent widely used to treat various tumors, bevacizumab has been approved to treat GBM in combination with TMZ and RT<sup>[31]</sup>, which can reduce glucocorticoid requirements to lower the risk of and other side effects caused by morbidity glucocorticoids. The use of bevacizumab is now the first-line treatment for a relapsed or progressing GBM,

slightly increasing patients' progression-free survival<sup>[32]</sup>.

Despite the standard therapeutic strategy used in GBM, the survival rate is still frustrating. Novel approaches are currently being explored, some of which have been approved for clinical trials. Inhibitor therapy, one of the high-profile molecular targeting treatments, typically targets a particular kinase or a group of kinases that are excessively activated in malignant tissues. Rearrangement, amplification, and fusion of receptor tyrosine kinase (RTK) are observed in GBM, making them the appealing targets for advanced treatment<sup>[33]</sup>. The EGFR inhibitor Afatinib, which is approved by the FDA to treat non-small-cell lung cancer, increases the overall survival of GBM patients when combined with TMZ<sup>[34]</sup>. Studies and trials for other EGFR inhibitors, including erlotinib and gefitinib, are also being conducted<sup>[35-36]</sup>. Besides RTK inhibitors, studies and trials for adenosine diphosphate (ADP) ribose polymerase inhibitors, myeloid cell leukemia-1, and topoisomerase are underway as well<sup>[33]</sup>.

The inbuilt immune system is the defense wall and strong weapon against pathogens and cancer cells. However, the immune system's ability to eliminate abnormal cells was suppressed under the tumor microenvironment. Immunotherapy has been proved to be effective in non-small cell lung cancer and quite a few other cancers via manipulating the related immune cells to rescue their ability to attack cancer cells. However, as immunologically cold tumors, gliomas are relatively insensitive to immunotherapy. The main forms of GBM immunotherapy under investigation include peptide vaccines, dendritic cell vaccines, chimeric T-cell receptors, checkpoint inhibitors, and oncolytic virotherapy<sup>[37]</sup>. Checkpoint inhibitors, such as anti-cytotoxic T-lymphocyteassociated protein 4 (CTLA-4) and anti-programmed cell death protein 1 (PD-1) drugs, appear to produce amphibolous outcomes. These may pioneer GBM immunotherapy through the conduct of numerous preclinical studies and clinical trials<sup>[38]</sup>.

The nano-therapy refers to the utilization of nanoparticulate anti-GBM drugs. Gold nanosphere, gold nanorods, carbon nanotubes, nanogels, polymeric nanoparticles, polymeric micelles, and liposomes are the main carriers of nanoparticulate anti-GBM drugs currently under investigation<sup>[33]</sup>. The unique characteristics of these novel materials result in easier diffusion through the blood-brain barrier, enhanced permeability and retention effect, and a homogenous

distribution within the tumor. With the application of these strategies, nano-formulated drugs, such as erlotinib, can be delivered through liposomal nanoparticles specifically to tumor cells<sup>[39–40]</sup>.

There are also some other new therapeutic strategies attempting to improve the prognosis of GBM. Tumor treating fields (TTF) therapy, also known as alternating electric field therapy, is a kind of non-invasive GBM treatment, which delivers lowintensity (1 to 3 V/cm), intermediate-frequency (100 to 300 kHz), alternating electric fields transcutaneously. The electric fields exert biophysical force on charged and polarizable molecules known as dipoles. TTF therapy has antimitotic effects and can interfere with DNA repair, thus potentially suppressing TMZ resistance mentioned earlier. Prevention of the inhibitory effects of the phosphatidylinositol 3-kinase (PI3K)/AKT/mammalian target of rapamycin (mTOR) signaling pathway on autophagy is another possible function of the TTF therapy in GBM treatment, because the PI3K pathway is closely related to GBM invasion. Antitumor immunity and anti-migratory through increasing cell membrane permeability are important anti-tumor mechanisms of the TTF therapy as well<sup>[41]</sup>.

Laser interstitial thermal therapy (LITT) is a neurosurgical technique utilizing thermal energy. Directed by the stereotaxic device, an optical fiber generates heat in the center of the tumor and burns tumor cells. The procedure only leaves a small hole on the skull and barely affects healthy tissues surrounding the tumor<sup>[33]</sup>. Prolonged survival of newly diagnosed GBM patients after LITT has been demonstrated<sup>[42]</sup>. For patients with unresectable tumors, the application of LITT is promising.

Stem cell-based therapy is the union of at least five types of cancer stem cell (CSC) targeted therapy, including chemoradiotherapy with radiosensitizers and chemotherapeutics, and immunotherapy mentioned above<sup>[43]</sup>. It is mainly based on the stem cell theory, in which CSCs in the brain tissue originate from stem cells with accumulated mutations, and the uncontrolled migration of CSCs finally causes tumorigenesis. A big merit of the stem cell-based therapy is the minimal side effects, but a multitude of issues remain to be addressed, for example, biosafety<sup>[44]</sup>.

Currently, systematic treatment of GBM includes surgery, radiation therapy (RT), and chemotherapy. Advanced therapies vary from research hotspots, such as immunotherapy, nano-therapy, and inhibitor

therapy, to TTF therapy and LITT (Fig. 1B). A traditional therapy is still the preferred choice when patients are first diagnosed with glioblastoma. Patients who receive chemotherapy or RT besides tumor resection have a better prognosis than those who do not<sup>[45]</sup>. Despite the application of improved protocols, i.e., the Stupp protocol, in which the doses of chemotherapeutic agents maintain at a low level, a recurrence is still almost a certainty due to drug or RT resistance. For progressing or recurrent GBM, additional advanced therapy is urgently needed. It is predictable that molecular targeted therapy and immune therapy will be added to the option list of GBM treatment in the future with the development of precision medicine. The combination of different therapies is also a future trend with more evidence supportive of its efficacy. The TTF therapy is already recommended in China, while clinical trials are needed for the addition of LITT to the standard treatment. To retard the ominous process of recurrence and increase survival, improvements in GBM treatment should be put on the top of the agenda.

## The patterns and dynamics of glioblastoma multiforme invasion

GBM cell invasion follows specific patterns. Although it can be attributed to the intrinsic genetic features of tumor cells or their interactions with the microenvironment, generally, there are several crucial signaling pathways supporting tumor cells cooperatively, including the p53 and the RTK pathways, as well as various pathways related to epithelial-mesenchymal transition (EMT). Growth factors, chemokines, and integrins are also involved in the process of metastasis and invasion<sup>[8]</sup>.

In the 1930s, German pathologist Hans Joachim Scherer first defined the routes for GBM invasion, which is the prelude to systematic research into the patterns and dynamics of GBM invasion. According to his observation, gliomas are localized in the white matter at first and can migrate along white matter structures, blood vessels, and ependymal surfaces, which is later referred to as the Scherer structure<sup>[46]</sup>. Tamura et al demonstrated that glioma stem cells (GSCs) possess a tendency to migrate towards the anterior corpus callosum from all directions, while the cortical area seems less likely to be the destination. This might be due to the complex and tight connections of neurons in these areas. These observations further proved Scherer's anticipation that GBM cells progress along paths with the least obstruction<sup>[47]</sup>.

GBM invasion starts from the migration of tumor cells located at the border, and usually, there is a leader cell, followed by other invading cells. An "invasive margin", defined by Alieva et al, is the protruding multicellular groups originating at the interphase between the tumor and the brain parenchyma. With the highest proportion of invading cells that move in a directed way, the invasive margin configuration becomes the most aggressive type of border. Another aggressive border is the "diffuse margin", which means individual cell migration into the invasive area of the brain parenchyma. Invading cells here possess high a velocity but are less directed. The "well-defined border" refers to tumor margins without protrusions. Cells here are also dynamic, but in a spread-free way. In fact, even cells at the tumor core are not static, and migratory cells can be found in all areas of the tumor, indicating that mobility was one of the intrinsic characteristics of GBM cells. Due to the influence of the microenvironment, however, only specific cells at the aforementioned specific borders invade<sup>[48]</sup>.

#### Signal pathways involved in invasion

Referred to as the "Guardian of the Genome"<sup>[49]</sup>, p53 is a transcriptional regulator that prevents damaged cells from invasion by integrating stress signals and promoting cell cycle arrest<sup>[50]</sup>. Alteration of p53 found in 25% to 30% of primary GBM and 60% to 70% of secondary GBM, is the most common molecular abnormality in GBM<sup>[51]</sup>. Missense mutations of TP53, deletions of cyclin dependent kinase inhibitor 2A (CDKN2A/ARF), and amplification of mouse double minute 2 (MDM2) are ordinary mutations of the p53 pathway<sup>[52]</sup> (Fig. 2). TP53 plays a pivotal role in the proliferation of CSCs.



*Fig.* 2 p53 pathway in glioblastoma multiforme invasion. The alteration of p53 can be mainly attributed to *TP53* mutation, *CDKN2A/ARF* deletion, and *MDM2* amplification, which leads to CSC proliferation, enabling the tumor to escape destruction and evolve into more invasive phenotypes. TP53: tumor protein p53; CDKN2A: cyclin dependent kinase inhibitor 2A; MDM2: mouse double minute 2; CSC: cancer stem cell.

With *TP53* mutations, CSCs are more likely to survive chemo-radiotherapy and thus leading to recurrence<sup>[53]</sup>. The *CDKN2A/ARF* locus is a frequently deregulated component of the p53 pathway. Retained ARF expression may generate more metastatic and invasive phenotypes of GBM<sup>[54]</sup>. MDM2, amplified in GBM, is an E3 protein ligase responsible for p53 degradation through a ubiquitin-dependent lysosome pathway<sup>[55]</sup>. Normally, p53 expression upregulates MDM2. However, mutant p53 cannot transactivate MDM2, so under the circumstances, it escapes destruction and causes various mutational effects, such as the overexpression of EGFR<sup>[51]</sup>. Inhibitors of MDM2/p53 interaction are under investigation and are promising in GBM treatment.

RTKs are a kind of membrane-spanning proteins functioning as receptors of cytoplasmic signaling effectors after phosphorylation. Fifty-eight RTKs divided into 20 classes have been detected in humans, and the abnormality of some specific classes, for example, EGFR, insulin receptor (INSR), vascular endothelial growth factor receptor (VEGFR), and fibroblast growth factor receptor (FGFR), is closely related to GBM proliferation, invasion, and drug resistance, leading to a poor prognosis<sup>[56]</sup>. The EGFR family is a group of RTKs comprising EGFR, human epidermal growth factor receptor-2 (ErbB2/HER2), ErbB3/HER3, and ErbB4, of which EGFR is involved in the RAS/RAF/mitogen-activated protein kinase kinase (MEK)/extracellular signal-regulated kinase (ERK) pathway, the PI3K/AKT pathway, the Janus kinase (JAK)/signal transducer and activator of transcription (STAT) pathway, and the protein kinase C (PKC) pathway, and may play a primary role in the GBM onset, resistance to therapy, and recurrence<sup>[56–57]</sup> (Fig. 3). Its amplification is commonly observed in GBM. Mutation, rearrangement, and altered splicing exist but are less frequent<sup>[58]</sup>. The RAS/RAF/MEK/ERK pathway is crucial to the regulation of cell proliferation, metabolism, survival, and apoptosis. Once transphosphorylated, EGFR binds to growth factor receptor bound protein 2 (GRB2) and recruits SHC, which next activates son of sevenless 1 (SOS1), a guanine nucleotide exchange factor, to



*Fig. 3* EGFR pathway in glioblastoma multiforme invasion. Of the EGFR pathway, the RAS/RAF/MEK/ERK pathway, the PI3K/AKT pathway, and the JAK/STAT3 pathway are the three most important ones. These pathways are closely related to GSC stemness, the inhibition of tumor cell apoptosis, and tumor progression. EGFR: epidermal growth factor receptor; MEK: mitogen-activated protein kinase kinase; ERK: extracellular regulated MAP kinase; PI3K: phosphatidylinositol 3-kinase; JAK: Janus kinase; STAT3: signal transducer and activator of transcription 3; GSC: glioma stem cell; FAK: focal adhesion kinase; GRB2: growth factor receptor bound protein 2; FOXO1: Forkhead Box O1; BAD: BCL2 associated agonist of cell death; TSC2: TSC complex submit 2; mTOR: mammalian target of rapamycin; HIF1: hypoxia-inducible factor 1; VEFG: vascular endothelial growth factor; NF-kB: nuclear factor kappa-B.

induce RAS to exchange guanosine triphosphate (GTP) to guanosine diphosphate (GDP). Activated RAS leads to RAF-1 phosphorylation, allowing it to bind to MEK1/2. Phosphorylated by MEK1/2, ERK1/2, with over one hundred downstream cytoplasmic and nuclear substrates, induces a variety of biological responses<sup>[59]</sup>. In respect of GBM invasion, for example, as a result of EGFR amplification, ERK is upregulated and stabilizes YTH N<sup>6</sup>-methyladenosine RNA binding protein 2 (YTHDF2), thus promoting tumor propagation possibly due to repression of target genes expression, including L-xylulose reductase (LXRA) and HIVEP zinc finger 2 (HIVEP2)<sup>[60]</sup>.

Another important pathway involved in GBM invasion is the PI3K/AKT pathway. There are three classes of PI3K, among which Class I PI3K is the downstream effector of EGFR. It can be recruited by the ErbB family or RAS and phosphorylates the membrane lipid phosphatidylinositol-4,5-bisphosphate phosphatidylinositol-3,4,5- $(PIP_2)$ to generate triphosphate (PIP<sub>3</sub>). PIP<sub>3</sub> can activate AKT, which promotes cell proliferation by inhibiting caspase-9 and phosphorylating the pro-apoptotic BCL2 associated agonist of cell death (BAD). The indirect activation of mTOR due to the phosphorylation of TSC complex subunit 2 (TSC2), an inhibitor of mTOR, as a result of AKT activation, can cause increased synthesis of cyclin D1, hypoxia-inducible factor 1 (HIF1) and vascular endothelial growth factor (VEGF), which can tumor progression. AKT promote can also phosphorylate MDM2 and indirectly cause ubiquitination of p53<sup>[59]</sup>. By increasing glucose uptake and regulating glycolytic enzymes, constitutivelyactivated AKT stimulates the glycolysis of tumor cells<sup>[61]</sup>. However, to meet the bioenergetic needs, GBM does not possess a fixed metabolic phenotype, and instead, it can switch between the glycolytic phenotype and oxidative phenotype, which makes it easily resistant to metabolic therapy targeting AKT and continue to progress<sup>[62]</sup>. More research needs to be done to develop therapies that target the metabolic status of GBM cells.

EGFR also leads to the activation of the protooncogene c-SRC, the product of the avian tumor virus Rous sarcoma virus.<sup>[59]</sup>. Src family kinases bind to focal adhesion kinase (FAK) in integrin-mediated cell adhesion. The FAK/Src complex then regulates cellular functions, such as survival, proliferation, migration, and invasion, via downstream signaling pathways<sup>[63]</sup>. FAK overexpression also directly stimulates the formation of invadopodia and promotes their activity by controlling the localization of Src<sup>[64]</sup>. The JAK/STAT pathway is another important oncogenic pathway related to EGFR, which promotes GBM cell proliferation, angiogenesis, resistance to apoptosis, and immune escape through downstream targets, such as Bcl-xL, Bcl-2l1, Bcl-2, cyclin D1, and c-Myc. JAK is the abbreviation for Janus kinase, while STAT is short for signal transducer and activator of transcription. STAT is a family of seven transcription factors, including STAT1, STAT2, STAT3, STAT4, STAT-5a, STAT-5b, and STAT6. Their activation can be attributable to several signaling pathways, including cytokines, non-RTKs, and EGFR mentioned above. Of the seven members in the STAT family, STAT3 is the most widely discussed for tumor proliferation. It is associated with Notch signaling, and the JAK/STAT3 can be indirectly activated by transforming growth factor- $\beta$ (TGF- $\beta$ ), which helps maintain the stemness of GSCs. TGF- $\beta$  can bind to nuclear factor kappa-B (NF- $\kappa$ B), elevating the activity and the ability of GSC selfrenewal<sup>[65]</sup>. The transcriptional regulation of GSCs largely contributes to Type 3 EMT, which will be discussed in the next part.

EMT is a very important process in organismal development, wound healing, and tissue fibrosis. During EMT, epithelial cells lose their junctions and apical-basal polarity, and the epithelial appearance changes into a spindle-shaped, mesenchymal morphology, which facilitates the invasion of the tumor with an increased cell motility<sup>[66]</sup>. The reversed process is called mesenchymal-epithelial transition (MET), which is needed to influence the metastatic competence at the site of recolonization<sup>[67]</sup>. EMT can be triggered by a range of stimuli, including hypoxia, alterations of metabolism, cytokines, growth factors, and anti-tumor drugs<sup>[68]</sup>. In particular, transcription factors zinc-finger E-box-binding (ZEB), the Snail family of zinc-finger transcription factors (SNAIL), SLUG (also a zinc-finger transcription factor), lymphoid enhancer factor (LEF), and the TWIST family have been proved to be related to EMT in GBM cell invasion via the repression of epithelial marker genes and activation of mesenchymal marker genes mediated by various signaling pathways. This leads to a decreased expression of E-cadherin, claudin, and occludins, which are essential to the detachment of tumor cells. To allow migration, proteases, such as matrix metalloproteinases (MMPs) and cathepsins, are produced to degrade the extracellular matrix (ECM) and form invadopodia<sup>[69–72]</sup>.

One of the major signaling pathways inducing EMT is the TGF- $\beta$  signaling pathway, which can be further divided into the mothers against decapentaplegic homologs (SMAD)-dependent signaling pathway (Fig. 4) and the SMAD-independent pathway. In the SMAD-dependent pathway, TGF-B phosphorylates SMAD2 and SMAD3, allowing them to bind to SMAD4 and translocate to the nucleus, which further induces the transcription of ZEB, SNAII and SANI2, LEF1, and TWIST<sup>[73]</sup>. With the activation of RAS and PI3K by TGF- $\beta$ , the SMAD-independent signaling intersects with RTK signaling pathways, including the RAS/RAF/MEK/ERK pathway, PI3K/AKT signaling cascade, and the JAK/STAT3 pathway<sup>[68]</sup>. The PI3K/AKT signaling cascade lifts the expression of Snail and Slug via the activation of NF- $\kappa$ B<sup>[74]</sup>, while the RAS/RAF/MEK/ERK pathway phosphorylates nucleus Jun proto-oncogene, AP-1 transcription factor subunit (c-Jun) to promote the expression of EMT related transcription factors<sup>[59,68]</sup>. Crosstalk between the Notch signaling and TGF-B signaling via the Notch intracellular domain (NICD) and SMAD2 can also initiate EMT. After Delta-like or Jagged family binds to the Notch receptor, NICD is cleaved by  $\gamma$ secretase and tumor necrosis factor-a-converting



*Fig. 4* **Pathways related to epithelial-mesenchymal transition.** Activation of the TGF- $\beta$  pathway leads to EMT through the regulation of SMAD and other downstream effectors. The WNT/ $\beta$ -catenin pathway intersects with the SMAD-dependent pathway and contributes to EMT cooperatively. TGF- $\beta$ : transforming growth factor- $\beta$ ; SMAD: mothers against decapentaplegic; WNT: Wingless; FZD: Frizzled; DVL: Dishevelled; LEF: lymphoid enhancer factor; ZEB: zinc-finger E-box-binding; MMP: matrix metalloproteinase; EMT: epithelial-mesenchymal transition.

enzyme (TACE), and then translocated to the nucleus, leading by nuclear localization motif, thus activating SNAIL<sup>[68,75]</sup>.

Another important signaling pathway in EMT is the Wingless (WNT)/ $\beta$ -catenin pathway (*Fig. 4*). WNT proteins recruit Dishevelled (DVL) after binding to the membrane receptor complex of Frizzled (FZD), which stabilizes  $\beta$ -catenin. Glycogen synthase kinase 3 (GSK3) further determines whether  $\beta$ -catenin is transported into the nucleus or phosphorylated and rapidly degrades. If transported, it binds to complementary transcription factors T-cell factor (TCF)/LEF complex, as well as stimulates ZEB, SLUG, and TWIST<sup>[76–77]</sup>.

The expression of the serine protease inhibitor (serpin) superfamily is reported to be mainly linked with the mesenchymal subtype GBM<sup>[78]</sup>. Accumulating evidence has revealed an important role that serpins play in EMT. The Serpin family H member 1 (SERPINH1) gene, which encodes Serpin H1, better known as heat shock protein 47 (HSP47), is reported to be involved in the activation of WNT/βcatenin pathway<sup>[79]</sup>. In addition, HSP47 overexpression promotes ECM-related genes mainly TGF-β signaling, which significantly through contributes tumor cell stemness to and tumorigenesis<sup>[80]</sup>. Serpin family Α member 3 expression also (SERPINA3) enhances GBM malignancy inducing cell stemness via and migration<sup>[81-82]</sup>. Other members of the serpin family, including SERPINE1 and SERPING1, have been proven to be positively related to GBM proliferation and progression<sup>[78,83]</sup>. A potential linkage between SERPINF1 or SERPINB9 and EMT has also been suggested<sup>[84-85]</sup>. Therefore, targeting SERPIN to prevent GBM invasion and recurrence might be feasible.

#### **Conclusions and future perspectives**

GBM, a malignant brain tumor, has both high occurrence and recurrence rates. A definite diagnosis procedure starts with imaging (usually MRI) and has to undergo histopathological examinations, in that a more appropriate targeted treatment can be settled upon. Traditional treatment includes maximal surgical resection of the tumor, radiation therapy, and chemotherapy. Advanced treatments include tumortreating fields therapy, immunotherapy, and stem cellbased therapy.

Despite all these treatment methods, current

outcomes are still unsatisfactory. The medium survival time of GBM patients, ranging from 11 months to two years, have not significantly improved over the past two decades<sup>[3,86]</sup>. Therapy resistance and recurrence are still inevitable. Specific anti-tumor drugs can even fasten GBM cell invasion by promoting EMT, which is such a paradox that brings about frustration to patients. This phenomenon might be the result of redundant compensatory mechanisms of GBM, insufficient target coverage, or poor tolerability and safety<sup>[87]</sup>.

Whatsoever, efforts are made to solve the dilemma. Different signaling pathways, either associated with inbuilt gene programs or related to the been microenvironment, have discovered and elucidated in recent years. Herein, we mainly discuss the p53 pathway, RTK pathways, and EMT-related pathways, especially their roles in promoting GBM invasion. These signaling pathways offer new insights into advanced GBM therapies. New drugs are to be designed to target elements involved in these pathways.

It is undeniable that humans are still far from the destination of effective GBM treatment. Nevertheless, by exploring possible targets and methods, we can anticipate that GBM patients will have more effective treatments in the future, and thus the financial burden can be largely relieved.

#### Fundings

This study was supported by National Natural Science Foundation of China (Grant No. 82003764 to L.F.) and Universities Natural Science Research Project of Jiangsu Province (Grant No. 19KJB350001 to L.F.).

#### Acknowledgments

None.

#### References

- Ostrom QT, Patil N, Cioffi G, et al. CBTRUS statistical report: primary brain and other central nervous system tumors diagnosed in the United States in 2013 –2017[J]. *Neuro Oncol*, 2020, 22(12 Suppl 2): iv1–iv96.
- [2] Tan AC, Ashley DM, López GY, et al. Management of glioblastoma: State of the art and future directions[J]. *CA A Cancer J Clin*, 2020, 70(4): 299–312.
- [3] Witthayanuwat S, Pesee M, Supaadirek C, et al. Survival

analysis of glioblastoma multiforme[J]. *Asian Pac J Cancer Prev*, 2018, 19(9): 2613–2617.

- [4] Zhu B, Wu X, Piao H, et al. A comparison of epidemiological characteristics of central nervous system Tumours in China and globally from 1990 to 2019[J]. *Neuroepidemiology*, 2021, 55(6): 460–472.
- [5] Ohgaki H, Kleihues P. The definition of primary and secondary glioblastoma[J]. *Clin Cancer Res*, 2013, 19(4): 764–772.
- [6] Wen J, Chen W, Zhu Y, et al. Clinical features associated with the efficacy of chemotherapy in patients with glioblastoma (GBM): a surveillance, epidemiology, and end results (SEER) analysis[J]. *BMC Cancer*, 2021, 21(1): 81.
- [7] Chouleur T, Tremblay ML, Bikfalvi A. Mechanisms of invasion in glioblastoma[J]. *Curr Opin Oncol*, 2020, 32(6): 631–639.
- [8] Vollmann-Zwerenz A, Leidgens V, Feliciello G, et al. Tumor cell invasion in glioblastoma[J]. *Int J Mol Sci*, 2020, 21(6): 1932.
- [9] Kanderi T, Gupta V. Glioblastoma Multiforme[M]. Treasure Island (FL): StatPearls Publishing, 2021.
- [10] Ruiz-López E, Calatayud-Pérez J, Castells-Yus I, et al. Diagnosis of glioblastoma by immuno-positron emission tomography[J]. *Cancers (Basel)*, 2021, 14(1): 74.
- [11] Romero-Rojas AE, Diaz-Perez JA, Amaro D, et al. Glioblastoma metastasis to parotid gland and neck lymph nodes: fine-needle aspiration cytology with histopathologic correlation[J]. *Head Neck Pathol*, 2013, 7(4): 409–415.
- [12] Jones J, Nguyen H, Drummond K, et al. Circulating biomarkers for Glioma: a review[J]. *Neurosurgery*, 2021, 88(3): E221–E230.
- [13] Wesseling P, Capper D. WHO 2016 Classification of gliomas[J]. *Neuropathol Appl Neurobiol*, 2018, 44(2): 139–150.
- [14] Mansouri A, Hachem LD, Mansouri S, et al. MGMT promoter methylation status testing to guide therapy for glioblastoma: refining the approach based on emerging evidence and current challenges[J]. *Neuro Oncol*, 2019, 21(2): 167–178.
- [15] Choi SW, Lee Y, Shin K, et al. Mutation-specific noncanonical pathway of PTEN as a distinct therapeutic target for glioblastoma[J]. *Cell Death Dis*, 2021, 12(4): 374.
- [16] Korshunov A, Schrimpf D, Ryzhova M, et al. H3-/IDH-wild type pediatric glioblastoma is comprised of molecularly and prognostically distinct subtypes with associated oncogenic drivers[J]. *Acta Neuropathol*, 2017, 134(3): 507–516.
- [17] Senhaji N, Louati S, Chbani L, et al. *EGFR* amplification and *IDH* mutations in glioblastoma patients of the Northeast of Morocco[J]. *Biomed Res Int*, 2017, 2017: 8045859.
- [18] Tejada S, Becerra-Castro MV, Nuñez-Cordoba J, et al. Ki-67 proliferative activity in the tumor margins as a robust prognosis factor in Glioblastoma patients[J]. *J Neurol Surg A Cent Eur Neurosurg*, 2021, 82(1): 53–58.

- [19] Suwala AK, Stichel D, Schrimpf D, et al. Glioblastomas with primitive neuronal component harbor a distinct methylation and copy-number profile with inactivation of TP53, PTEN, and RB1[J]. *Acta Neuropathol*, 2021, 142(1): 179–189.
- [20] Koruga N, Pekmezović T, Tomaš I, et al. Factors affecting outcome in the treatment of glioblastoma[J]. *Acta Clin Croat*, 2022, 60(3): 373–378.
- [21] Montemurro N, Fanelli GN, Scatena C, et al. Surgical outcome and molecular pattern characterization of recurrent glioblastoma multiforme: A single-center retrospective series[J]. *Clin Neurol Neurosurg*, 2021, 207: 106735.
- [22] Barani IJ, Larson DA. Radiation therapy of glioblastoma[M]//Raizer J, Parsa A. Current Understanding and Treatment of Gliomas. Cham: Springer, 2015: 49–73.
- [23] Vanhove C, Goethals I. Magnetic resonance imaging-guided radiation therapy using animal models of glioblastoma[J]. Br J Radiol, 2019, 92(1095): 20180713.
- [24] Antoni D, Feuvret L, Biau J, et al. Radiation guidelines for gliomas[J]. *Cancer Radiothér*, 2022, 26(1–2): 116–128.
- [25] Li R, Wang H, Liang Q, et al. Radiotherapy for glioblastoma: clinical issues and nanotechnology strategies[J]. *Biomater Sci*, 2022, 10(4): 892–908.
- [26] Zhang Y, Dai R, Chen Z, et al. Efficacy and safety of carmustine wafers in the treatment of glioblastoma multiforme: a systematic review[J]. *Turk Neurosurg*, 2014, 24(5): 639–645.
- [27] Anjum K, Shagufta BI, Abbas SQ, et al. Current status and future therapeutic perspectives of glioblastoma multiforme (GBM) therapy: A review[J]. *Biomed Pharmacother*, 2017, 92: 681–689.
- [28] Rajaratnam V, Islam MM, Yang M, et al. Glioblastoma: Pathogenesis and current status of chemotherapy and other novel treatments[J]. *Cancers (Basel)*, 2020, 12(4): 937.
- [29] LiverTox: Clinical and research information on drug-induced liver Injury[M]. Bethesda (MD): National Institute of Diabetes and Digestive and Kidney Diseases, 2012.
- [30] Jiapaer S, Furuta T, Tanaka S, et al. Potential strategies overcoming the temozolomide resistance for glioblastoma[J]. *Neurol Med Chir (Tokyo)*, 2018, 58(10): 405–421.
- [31] Chinot OL, Wick W, Mason W, et al. Bevacizumab plus radiotherapy-temozolomide for newly diagnosed glioblastoma[J]. *N Engl J Med*, 2014, 370(8): 709–722.
- [32] Garcia J, Hurwitz HI, Sandler AB, et al. Bevacizumab (Avastin<sup>®</sup>) in cancer treatment: A review of 15 years of clinical experience and future outlook[J]. *Cancer Treat Rev*, 2020, 86: 102017.
- [33] Wu W, Klockow JL, Zhang M, et al. Glioblastoma multiforme (GBM): An overview of current therapies and mechanisms of resistance[J]. *Pharmacol Res*, 2021, 171: 105780.
- [34] Vengoji R, Macha MA, Nimmakayala RK, et al. Afatinib and Temozolomide combination inhibits tumorigenesis by targeting EGFRvIII-cMet signaling in glioblastoma cells[J]. J

Exp Clin Cancer Res, 2019, 38(1): 266.

- [35] Bagherian A, Roudi B, Masoudian N, et al. Antiglioblastoma effects of nanomicelle-curcumin plus erlotinib[J]. *Food Funct*, 2021, 12(21): 10926–10937.
- [36] Blandin AF, Da Silva EC, Mercier MC, et al. Gefitinib induces EGFR and  $\alpha 5\beta 1$  integrin co-endocytosis in glioblastoma cells[J]. *Cell Mol Life Sci*, 2021, 78(6): 2949–2962.
- [37] Huang B, Li X, Li Y, et al. Current immunotherapies for glioblastoma multiforme[J]. *Front Immunol*, 2020, 11: 603911.
- [38] De Felice F, Pranno N, Marampon F, et al. Immune checkpoint in glioblastoma multiforme[J]. *Crit Rev Oncol Hematol*, 2019, 138: 60–69.
- [39] Alphandéry E. Nano-therapies for glioblastoma treatment[J]. Cancers (Basel), 2020, 12(1): 242.
- [40] Lakkadwala S, Singh J. Co-delivery of doxorubicin and erlotinib through liposomal nanoparticles for glioblastoma tumor regression using an *in vitro* brain tumor model[J]. *Colloids Surf B Biointerfaces*, 2019, 173: 27–35.
- [41] Rominiyi O, Vanderlinden A, Clenton SJ, et al. Tumour treating fields therapy for glioblastoma: current advances and future directions[J]. *Br J Cancer*, 2021, 124(4): 697–709.
- [42] Khan AB, Matsuoka CK, Lee S, et al. Prolonged survival after laser interstitial thermal therapy in glioblastoma[J]. *Surg Neurol Int*, 2021, 12: 228.
- [43] Zhang Q, Xiang W, Yi D, et al. Current status and potential challenges of mesenchymal stem cell-based therapy for malignant gliomas[J]. *Stem Cell Res Ther*, 2018, 9(1): 228.
- [44] Abadi B, Ahmadi-Zeidabadi M, Dini L, et al. Stem cell-based therapy treating glioblastoma multiforme[J]. *Hematol Oncol Stem Cell Ther*, 2021, 14(1): 1–15.
- [45] Sun K, Zhou X, Li T, et al. Clinicopathological characteristics and treatment outcomes of epithelioid glioblastoma[J]. *Neurosurg Rev*, 2021, 44(6): 3335–3348.
- [46] Reuter G, Lombard A, Suero ME, et al. Hans Joachim Scherer: an under-recognized pioneer of glioma research in Belgium[J]. *Acta Neurol Belg*, 2021, 121(4): 867–872.
- [47] Tamura R, Miyoshi H, Sampetrean O, et al. Visualization of spatiotemporal dynamics of human glioma stem cell invasion[J]. *Mol Brain*, 2019, 12(1): 45.
- [48] Alieva M, Leidgens V, Riemenschneider MJ, et al. Intravital imaging of glioma border morphology reveals distinctive cellular dynamics and contribution to tumor cell invasion[J]. *Sci Rep*, 2019, 9(1): 2054.
- [49] Lane DP. p53, guardian of the genome[J]. *Nature*, 1992, 358(6381): 15–16.
- [50] Kastenhuber ER, Lowe SW. Putting p53 in context[J]. Cell, 2017, 170(6): 1062–1078.
- [51] England B, Huang T, Karsy M. Current understanding of the role and targeting of tumor suppressor p53 in glioblastoma multiforme[J]. *Tumor Biol*, 2013, 34(4): 2063–2074.
- [52] Zhang Y, Dube C, Gibert M Jr, et al. The p53 pathway in

glioblastoma[J]. Cancers (Basel), 2018, 10(9): 297.

- [53] Orzan F, De Bacco F, Crisafulli G, et al. Genetic evolution of glioblastoma stem-like cells from primary to recurrent tumor[J]. *Stem Cells*, 2017, 35(11): 2218–2228.
- [54] Fontana R, Ranieri M, La Mantia G, et al. Dual role of the alternative reading frame ARF protein in cancer[J]. *Biomolecules*, 2019, 9(3): 87.
- [55] Stark AM, Witzel P, Strege RJ, et al. p53, mdm2, EGFR, and msh2 expression in paired initial and recurrent glioblastoma multiforme[J]. *J Neurol Neurosurg Psychiatry*, 2003, 74(6): 779–783.
- [56] Tilak M, Holborn J, New LA, et al. Receptor tyrosine kinase signaling and targeting in glioblastoma multiforme[J]. *Int J Mol Sci*, 2021, 22(4): 1831.
- [57] An Z, Aksoy O, Zheng T, et al. Epidermal growth factor receptor and EGFRvIII in glioblastoma: signaling pathways and targeted therapies[J]. *Oncogene*, 2018, 37(12): 1561–1575.
- [58] Eskilsson E, Røsland GV, Solecki G, et al. EGFR heterogeneity and implications for therapeutic intervention in glioblastoma[J]. *Neuro Oncol*, 2018, 20(6): 743–752.
- [59] Wee P, Wang Z. Epidermal growth factor receptor cell proliferation signaling pathways[J]. *Cancers (Basel)*, 2017, 9(5): 52.
- [60] Fang R, Chen X, Zhang S, et al. EGFR/SRC/ERK-stabilized YTHDF2 promotes cholesterol dysregulation and invasive growth of glioblastoma[J]. *Nat Commun*, 2021, 12(1): 177.
- [61] Hosios AM, Manning BD. Cancer signaling drives cancer metabolism: AKT and the warburg effect[J]. *Cancer Res*, 2021, 81(19): 4896–4898.
- [62] Duraj T, García-Romero N, Carrión-Navarro J, et al. Beyond the Warburg effect: oxidative and glycolytic phenotypes coexist within the metabolic heterogeneity of glioblastoma[J]. *Cells*, 2021, 10(2): 202.
- [63] Dong C, Li X, Yang J, et al. PPFIBP1 induces glioma cell migration and invasion through FAK/Src/JNK signaling pathway[J]. *Cell Death Dis*, 2021, 12(9): 827.
- [64] Mousson A, Legrand M, Steffan T, et al. Inhibiting FAKpaxillin interaction reduces migration and invadopodiamediated matrix degradation in metastatic melanoma cells[J]. *Cancers (Basel)*, 2021, 13(8): 1871.
- [65] Ou A, Ott M, Fang D, et al. The Role and Therapeutic Targeting of JAK/STAT Signaling in Glioblastoma[J]. *Cancers (Basel)*, 2021, 13(3): 437.
- [66] Dongre A, Weinberg RA. New insights into the mechanisms of epithelial-mesenchymal transition and implications for cancer[J]. *Nat Rev Mol Cell Biol*, 2019, 20(2): 69–84.
- [67] Zavadil J, Haley J, Kalluri R, et al. Epithelial-mesenchymal transition[J]. *Cancer Res*, 2008, 68(23): 9574–9577.
- [68] Majc B, Sever T, Zarić M, et al. Epithelial-to-mesenchymal transition as the driver of changing carcinoma and glioblastoma microenvironment[J]. *Biochim Biophys Acta* (BBA) Mol Cell Res, 2020, 1867(10): 118782.

- [69] Chow KH, Park HJ, George J, et al. S100A4 is a biomarker and regulator of Glioma stem cells that is critical for mesenchymal transition in glioblastoma[J]. *Cancer Res*, 2017, 77(19): 5360–5373.
- [70] Wirsik NM, Ehlers J, Mäder L, et al. TGF-β activates pericytes via induction of the epithelial-to-mesenchymal transition protein SLUG in glioblastoma[J]. *Neuropathol Appl Neurobiol*, 2021, 47(6): 768–780.
- [71] Du L, Lee JH, Jiang H, et al. β-Catenin induces transcriptional expression of PD-L1 to promote glioblastoma immune evasion[J]. *J Exp Med*, 2020, 217(11): e20191115.
- [72] Yang W, Wu P, Ma J, et al. Sortilin promotes glioblastoma invasion and mesenchymal transition through GSK-3β/βcatenin/twist pathway[J]. *Cell Death Dis*, 2019, 10(3): 208.
- [73] Gonzalez DM, Medici D. Signaling mechanisms of the epithelial-mesenchymal transition[J]. *Sci Signal*, 2014, 7(344): re8.
- [74] Rajendran P, Ben Ammar R, Al-Saeedi FJ, et al. Thidiazuron decreases epithelial-mesenchymal transition activity through the NF-kB and PI3K/AKT signalling pathways in breast cancer[J]. J Cell Mol Med, 2020, 24(24): 14525–14538.
- [75] Deshmukh AP, Vasaikar SV, Tomczak K, et al. Identification of EMT signaling cross-talk and gene regulatory networks by single-cell RNA sequencing[J]. *Proc Natl Acad Sci U S A*, 2021, 118(19): e2102050118.
- [76] Coelho BP, Fernandes C, Boccacino JM, et al. Multifaceted WNT signaling at the crossroads between epithelialmesenchymal transition and autophagy in glioblastoma[J]. *Front Oncol*, 2020, 10: 597743.
- [77] Sha Z, Zhou J, Wu Y, et al. BYSL promotes glioblastoma cell migration, invasion, and mesenchymal transition through the GSK-3β/β-catenin signaling pathway[J]. *Front Oncol*, 2020, 10: 565225.
- [78] Jovčevska I. Genetic secrets of long-term glioblastoma survivors[J]. Bosn J Basic Med Sci, 2019, 19(2): 116–124.
- [79] Tian S, Peng P, Li J, et al. SERPINH1 regulates EMT and gastric cancer metastasis via the Wnt/β-catenin signaling pathway[J]. *Aging (Albany NY)*, 2020, 12(4): 3574–3593.
- [80] Jiang X, Zhou T, Wang Z, et al. HSP47 Promotes glioblastoma stemlike cell survival by modulating tumor microenvironment extracellular matrix through TGF-β pathway[J]. ACS Chem Neurosci, 2017, 8(1): 128–134.
- [81] Norton ES, Da Mesquita S, Guerrero-Cazares H. SERPINA3 in glioblastoma and Alzheimer's disease[J]. *Aging (Albany NY)*, 2021, 13(18): 21812–21813.
- [82] Nimbalkar VP, Kruthika BS, Sravya P, et al. Differential gene expression in peritumoral brain zone of glioblastoma: role of SERPINA3 in promoting invasion, stemness and radioresistance of glioma cells and association with poor patient prognosis and recurrence[J]. *J Neurooncol*, 2021, 152(1): 55–65.

- [83] Ma T, Qian K, Xu T, et al. Serpin peptidase inhibitor, clade E nexin group 1 promotes cellular proliferative capacities and malignant behaviors in glioblastoma through upregulating hairy and enhancer of split-1[J]. *J BUON*, 2021, 26(5): 2074–2083.
- [84] Polisetty RV, Gupta MK, Nair SC, et al. Glioblastoma cell secretome: analysis of three glioblastoma cell lines reveal 148 non-redundant proteins[J]. *J Proteomics*, 2011, 74(10): 1918–1925.
- [85] Jiang H, Jin C, Liu J, et al. Next generation sequencing

analysis of miRNAs: MiR-127 –3p inhibits glioblastoma proliferation and activates TGF- $\beta$  signaling by targeting SKI[J]. *OMICS*, 2014, 18(3): 196–206.

- [86] Lacroix M, Abi-Said D, Fourney DR, et al. A multivariate analysis of 416 patients with glioblastoma multiforme: prognosis, extent of resection, and survival[J]. *J Neurosurg*, 2001, 95(2): 190–198.
- [87] Le Rhun E, Preusser M, Roth P, et al. Molecular targeted therapy of glioblastoma[J]. *Cancer Treat Rev*, 2019, 80: 101896.

#### CLINICAL TRIAL REGISTRATION

The *Journal* requires investigators to register their clinical trials in a public trials registry for publication of reports of clinical trials in the *Journal*. Information on requirements and acceptable registries is available at https://clinicaltrials.gov/.